

## RESEARCH OF ANTIOXIDANT AND PROOXIDANT PROPERTIES OF GLYCYRRHETIC ACID DERIVATIVES

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**Abstract.** *It was studied the effect of new synthetic derivatives of glycyrrhetic acid (GRA) on the functional mitochondrial parameters, as well as the parameters of the CsA-sensitive pore of mitochondrial membranes in the article. For the first time it was shown that GRA derivatives 2-(N-cytisine)-ethyl-3-0-acetyl-18βH-glycyrrhetate (cytisine-ethyl-HC), 2-(N-cytisine)-isopropyl-3-0-acetyl-18βH-glycyrrhetate (cytisine-isopropyl-HC) and N-(2-pyridyl)-3-0-acetyl-11-ketoolean-12-en-30-amide (2-pyridyl-HC-amide) inhibited the activity of the CsA-sensitive pore and have a protective effect on mitochondrial membranes. At the same time, the damaging effect of Ca<sup>2+</sup> ions and the LPO process decreases. Other GRA derivatives are N-(4-pyridyl)-3-0-acetyl-11-ketoolean-12-en-30-amide (4-pyridyl-HC-amide) and 2-(N-morpholine)-ethyl-3-0-acetyl-18βH-glycyrrhetate (morpholine-HC) uncouples OXPHOS, enhances the damaging effect of inducers of the CsA-sensitive pore on membranes, increasing their permeability to cations*

**Keywords:** *mitochondria, membrane permeability, CsA-sensitive pore, lipid peroxidation, oxidative phosphorylation (OXPHOS), antioxidants, prooxidants, free radicals, apoptosis, necrosis, glycyrrhetic acid derivatives.*

### Introduction

Modern scientific data suggest that mitochondria and the CsA-sensitive pore are targets for the action of various biologically active substances, pathogens, and pharmaceuticals (Kamburova, 2001; Akinshina, 2001; Dehpour et al., 1999; Чулиев, И.Н и.др, 2005). In this regard, the mechanisms of regulation of the functional state of the Ca<sup>2+</sup>-dependent CsA-sensitive mitochondrial pore and other Ca<sup>2+</sup>-dependent intracellular processes by biologically active compounds have recently been actively studied in many laboratories around the world (Zeuzem, 1998). To regulate the functional parameters and state of the CsA-sensitive pore, herbal preparations are often used, the biological and pharmacological activity of which is due to their membrane-active properties. Chemical modification of natural compounds can change their biological and pharmacological properties (Baltina et al., 1992; Tolstikov et al., 1997; Beskina et al., 2000; Beskina, 2002; Kamburova, 2001).

The study of the role of lipid peroxidation (LPO) in the regulation of the most important functions of the cell is of interest for a number of reasons (Vladimirov, 2000). LPO induction in mitochondria leads to a change in membrane permeability, a decrease in membrane potential, uncoupling of OXPHOS and ATP hydrolysis. The effect of LPO on mitochondrial functions is realized both at the level of the direct effect of LPO products on the lipid matrix of membranes, and various indirect effects.

One of the most important mechanisms through which LPO reactions can indirectly regulate mitochondrial functions is the CsA-sensitive pore, the transition of which to the open state is considered as an essential stage of mitochondrial damage during oxidative stress (OS) and associated necrosis or apoptosis. Free radicals are highly reactive compounds that can damage the structure and function of animal and plant cells (Sattorova, I. Y., & Cho'liyev, I. N., 2020). Organisms are constantly exposed to them. First, they are constantly formed as a result of natural metabolic processes occurring in the cell. Secondly, free radicals are formed under the influence of external factors, both natural and anthropogenic or man-made (under the influence of polluted environment, smoking, radiation, household chemicals).

It is known that plant compounds are the main source of biological material in the production of drugs with antioxidant properties (Potapovich., 2003; Tolstikov et al., 1997).

It is known that free radicals are the main cause of many human and animal diseases (Vladimirov, 1998). The body also has an antioxidant system that protects the body from free radicals. Antioxidants are able to neutralize the activity of free radicals, protect cell membrane phospholipids from oxidation (Vladimirov, 1998).

A natural question arises as to whether the glycyrrhetic acid derivatives that transform the CsA-sensitive pore into a closed configuration have antioxidant properties. In this regard, we studied the effect of glycyrrhetic acid and its derivatives on the LPO process of mitochondrial membranes. Cumene hydroperoxide (CHP) was used as an LPO inducer.

### **Results and discussion**

As a result of the studies, it was found that glycyrrhetic acid increases the accumulation of malondialdehyde (MDA) in mitochondrial membranes by 40% (Fig. 1). Similar data were also obtained by other authors. The addition of other derivatives of glycyrrhetic acid - 2-pyridyl-HC-amide, cytosine-isopropyl-HC and cytosine-ethyl-HC at a concentration of 50  $\mu\text{M}$  prevented the effect of CHP on the level of MDA in isolated liver mitochondria. At the same time, the decrease in the accumulation of MDA was 20%, 40% and 45%, respectively, relative to the control (Fig. 1).

Subsequently, we studied the effect of GrK derivatives on the LPO system induced by  $\text{Fe}^{2+}$ -ascorbate (Fig. 2). Under these conditions, the derivatives of glycyrrhetic acid - 2-pyridyl-HC-amide, cytosine-isopropyl-HC and cytosine-ethyl-HC at a concentration of 50  $\mu\text{M}$  prevented the effect of  $\text{Fe}^{2+}$ -ascorbate on the level of MDA in isolated mitochondria by 15%, 30.6% and 50 %, respectively.

Thus, we found that glycyrrhetic acid derivatives: 2-pyridyl-HC-amide, cytosine-isopropyl-HC and cytosine-ethyl-HC have antioxidant properties and have a protective effect on mitochondria, reducing the damaging effect of CHP and the LPO process.

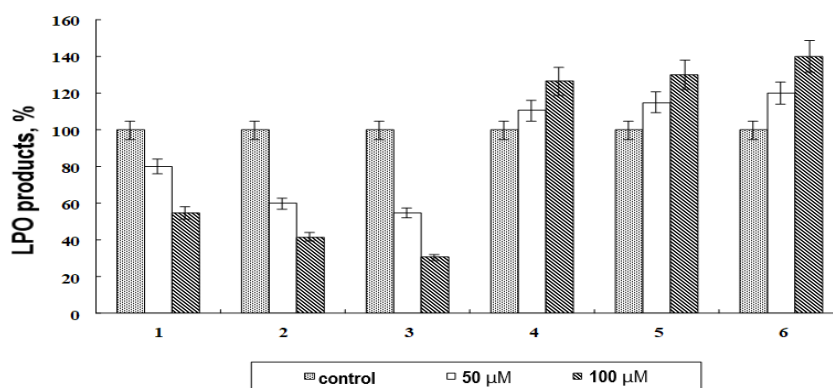
One of the mechanisms through which LPO reactions can indirectly regulate mitochondrial functions is the CsA-sensitive pore. Possibly, this mechanism is influenced by biologically active compounds: 2-pyridyl-HC-amide, cytosine-isopropyl-HC and cytosine-ethyl-HC on the functions of mitochondria.

We also studied the effect of other derivatives of glycyrrhetic acid on the LPO process of mitochondrial membranes, using LPO inductors - CHP and the  $\text{Fe}^{2+}$ -ascorbate system. When studying the effect of morpholine-HC, 4-pyridyl-HC-amide on the lipid peroxidation of mitochondrial membranes, it was shown that these derivatives acted on the state of the mitochondrial pore, increasing the passive permeability of the membranes and reducing the  $\text{Ca}^{2+}$ -

capacity of mitochondria. Previous experiments with glycyrrhetic acid showed that it increases the accumulation of MDA in mitochondrial membranes. However, the effects of the compounds of morpholine-HC and 4-pyridyl-HC-amide at a concentration of 50  $\mu\text{M}$  are weaker than the classical LPO inducers - CHP and the  $\text{Fe}^{2+}$ -ascorbate system (Fig. 1., Fig. 2 and Fig. 3). An increase in the concentration of drugs in the incubation medium led to a further increase in the accumulation of MDA in mitochondrial membranes.

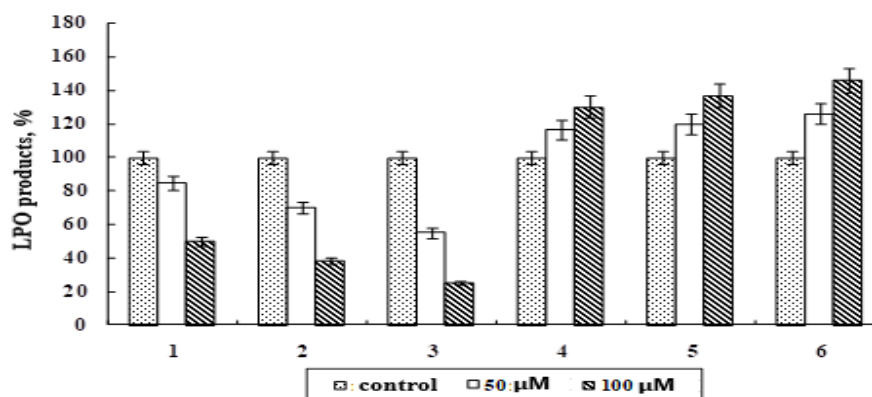
As experiments have shown, in the presence of CHP, an increase in the accumulation of MDA is observed. Against this background, preparations of morpholine-HC and 4-pyridyl-HC-amide (50  $\mu\text{M}$ ) led to a further increase in the accumulation of MDA in mitochondrial membranes. Higher concentrations of morpholine-HC and 4-pyridyl-HC-amide in SI led to a further increase in MDA accumulation in mitochondrial membranes by 26% and 30%, respectively. The results obtained confirm our assumption that the morpholine-HC and 4-pyridyl-HC-amide compounds have prooxidant properties (Fig. 1.).

Similar results were also obtained during induction by the  $\text{Fe}^{2+}$ -ascorbate system (Fig. 2). Subsequently, we studied the effect of morpholine-HC and 4-pyridyl-HC-amide on the LPO system induced by  $\text{Fe}^{2+}$ -ascorbate (Fig. 2 and Fig. 3). Under the same conditions, the preparations tested by us at a concentration of 50  $\mu\text{M}$  contributed to a further increase in the accumulation of MDA in mitochondrial membranes by 30% and 37%, respectively.



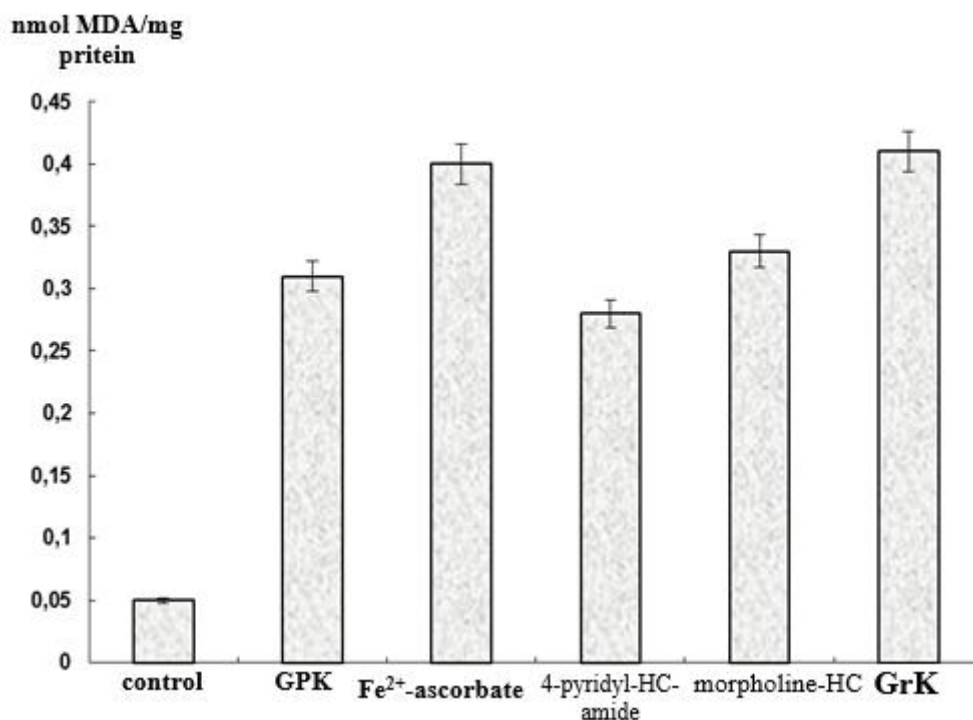
**Fig. 1. Effect of glycyrrhetic acid derivatives on GPA-dependent LPO**

1. - 2-pyridyl-GC-amide; 2. - cytosine-isopropyl-HC; 3. - cytosine-ethyl-HC; 4. - 4-pyridyl- GC - amide; 5. - morpholine- GC; 6. - glycyrrhetic acid. (n=6, P<0.05).



**Fig. 2. Effect of glycyrrhetic acid derivatives on Fe-ascorbate dependent LPO**

1. - 2-pyridyl-HC-amide; 2. - cytosine-isopropyl-HC; 3. - cytosine-ethyl-HC; 4. - 4-pyridyl-HC - amide; 5. - morpholine-HC; 6. - GrK. (n=6, P<0.05).



**Fig. 3. Influence of GrK derivatives - GrK morpholine-HC and 4-pyridyl-HC-amide on the LPO system\***

**Notes.** The concentration of GrA and its derivatives is 50  $\mu$ M. (n =6, P<0,05).

It is known that one of the mechanisms of violation of the CsA-sensitive pore is the intensification of lipid peroxidation against the background of a decrease in the activity of antioxidant defense enzymes - catalase and superoxide dismutase. As a result, membrane desensitization and an increase in their permeability to various ions and substances are observed. Some of the compounds we studied reduced the level of MDA in membranes that indicated their antioxidant properties.

Thus, 2-pyridyl-HC-amide, cytosine-isopropyl-HC and cytosine-ethyl-HC have antioxidant properties and have a protective effect on mitochondria, reducing the damaging effect of CHP and the Fe<sup>2+</sup>-ascorbate system, and other glycyrrhetic acid derivatives: morpholine-HC and 4-pyridyl-HC-amide have pro-oxidant properties, enhancing the damaging effect of CHP and the Fe<sup>2+</sup>-ascorbate system.

## REFERENCES

1. Akinshina N.G. Bioenergetic disorders in liver mitochondria during intoxication and possible methods of correction: Abstract of the thesis. diss. ... cand. biol. Sciences. - T.: 2001. - 24 s.
2. Khan R, Khan AQ, Lateef A et al. Glycyrrhizic acid suppresses the development of precancerous lesions via regulating the hyperproliferation, inflammation, angiogenesis and apoptosis in the colon of Wistar rats. PLoS One. 2013;8(2):e56020.
3. Elsherbini AM, Maysarah NM, El-Sherbiny et al. Glycyrrhizic acid ameliorates sodium nitrite-induced lung and salivary gland toxicity: Impact on oxidative stress, inflammation and fibrosis. Hum Exp Toxicol. 2021 Apr;40(4):707-721.
4. Beskina O.A. Novyye aspekty mekhanizma deystviya glitsirrizinovoy kisloty: Dis. ... kand. biol. nauk. - Tashkent, 2002. - 23 s.
5. Boldyrev A.A. Vvedeniye v biomembranologiyu. - Moskva.: Universitet, 1990. - S.77-78.

6. Vladimirov YU.A. Biologicheskiye membrany i nezaprogrammirovannaya smert' kletki // Sorosovskiy obrazovatel'nyy zhurnal. – 2000. – № 6 (9). – С. 2–9.
7. Vladimirov YU.A. Svobodnyye radikaly i antioksidanty. // Vestnik RAMN. - 1998. - №8. - S.43-51.
8. Kamburova V.S. Regulyatsiya tsiklosporin A – chuvstvitel'noy pory mitokhondriy: efekty glitsirrizinovoy kisloty i yeye aglikona: Diss. ... kand. biol. nauk. T.: 2001. – 110 s.
9. Fouladi S, Masjedi M, Ganjalikhani Hakemi M, Eskandari N. The Review of in Vitro and in Vivo Studies over the Glycyrrhizic Acid as Natural Remedy Option for Treatment of Allergic Asthma. *Iran J Allergy Asthma Immunol.* 2019 Feb;18(1):1-11.
10. Kicinska A, Jarmuszkiewicz W. Flavonoids and Mitochondria: Activation of Cytoprotective Pathways? *Molecules.* 2020 Jul 4;25(13):3060
11. Dehpour A.R., Zahedi H., Amini Sh., Akhgari M., Abdollahi M. Effects of glycyrrhiza derivatives against acetaminophen-induced hepatotoxicity. // *Iran J Med Sci.* – 1999. - V.24(1&2). – P.26-31.
12. Kao TC, Wu CH, Yen GC. Glycyrrhizic acid and 18 $\beta$ -glycyrrhetic acid recover glucocorticoid resistance via PI3K-induced AP1, CRE and NFAT activation. *Phytomedicine.* 2013 Feb 15;20(3-4):295-302.
13. Jain R, Hussein MA, Pierce S et al. Oncopreventive and oncotherapeutic potential of licorice triterpenoid compound glycyrrhizin and its derivatives: Molecular insights. *Pharmacol Res.* 2022 Apr;178:106138.
14. Чулиев, И. Н., & Камбурова В.С, Д. А. (2005). 2 (М-цитизин) этил-3-0-ацетил-18-3-Н-глицерретат как модификатор мегаканала митохондрий. *ДАН РУз*, (1), 41-47.
15. Sattorova, I. Y., & Cho'liyev, I. N. (2022). APOPTOSIS AND NECROSIS-PROGRAMMED CELL DEATH AND ITS ROLE IN CELL LIFE ACTIVITY. *Spectrum Journal of Innovation, Reforms and Development*, 3, 125-131.